Cell cycle inhibitors in oral leukoplakia: A review

Authors:

1) Dr. Gokul Sridharan MDS Reader,

Dept. of Oral Pathology,
YMT Dental College and Hospital,
Institutional area, Sector-4,
Kharghar,
Navi Mumbai- 410210
E-mail: drgokuls@gmail.com

2) Dr. Kriti Bagri-Manjrekar MDS Reader,

Dept. of Oral Pathology, YMT Dental College and Hospital, Institutional area, Sector-4, Kharghar, Navi Mumbai- 410210 E-mail: drkritibagri@gmail.com

3) Dr. Akhil Shankar MDS Reader,

Vistaa Dental Care, Mumbai, India E-mail: <u>akhil1904@gmail.com</u>

Corresponding author: Dr. Gokul Sridharan MDS, Reader,

Dept. of Oral Pathology, YMT Dental College and Hospital, Institutional area, Sector-4, Kharghar, Navi Mumbai- 410210

Navi Mumbai- 410210 Email: drgokuls@gmail.com

Conflict of interest: None

Source of grants: None

Key words: leukoplakia, cell cycle inhibitors, malignant transformation

Abstract:

Cell cycle is a complex and coordinated process characterized by orderly progression in the life cycle of cells orchestrated by cyclins and cyclin dependent kinases (CDK) and their inhibitors. The kinases drive the cell cycle by phosphorylating critical target proteins that are required for progression of cell to the next phase of cell cycle. This progress is tightly regulated by a group of cell cycle inhibitors known as CDK inhibitors which may be responsible for maintenance of cell cycle and proliferation arrest once the cells' developmental fate has been reached. Cell cycle inhibitors may act as checkpoints to identify the defective components of the cell thereby aiding in its repair and hence inactivation of these genes may interfere with the terminal differentiation leading to unrestricted proliferation and tumorigenesis. Cell proliferation is an important property of malignant tumor cells and dysregulation of genes governing the cell cycle is of considerable significance in the development of oral squamous cell carcinoma. Most OSCC generally develop by the malignant transformation of potentially malignant disorders the most common of which is leukoplakia. Hence evaluation of these genetic alterations in leukoplakia can be useful in assessment of the potential for malignant transformation and may also aid in attempting any therapeutic strategies at this stage. This review aims to provide data related to the role of cell cycle inhibitors in oral leukoplakia highlighting their role in malignant transformation.

1. Introduction

Cell division consists of two consecutive processes mainly consisting of DNA replication and segregation of daughter chromosomes into two separate cells. Replication of DNA occurs in a specific part of the interphase known as S phase (Vermeulen et al 2003). The transition from one cell cycle phase to another occurs in an orderly fashion and is regulated by cellular proteins. The cycle molecular principles of cell regulation have been defined largely in yeast, and are applicable in human beings which points to a number of surveillance check points that monitor the cell cycle and halt its progression (Cheng 2004). In mammalian cells, the cell cycle machinery which regulates the cell cycle operates mainly in the G1 phase (Cheng 2004). The key regulatory proteins of the cell cycle include the cyclin-dependent kinases (CDK's) which belong to the family of serine/threonine protein kinases that are activated at specific points of cell cycle (Vermeulen et al 2003). The CDK's join the regulatory proteins called cyclins to drive the cell through the complete cycle (Dickson & Schwartz 2009). The pattern of cyclin expression defines the cell progression and there are at least nine CDK's (CDK 1-9) and many cyclins which interact upon activation by specific phosphorylation (Kaldis et al 1998). While CDK's are positively regulated by cyclins through phosphorylation mechanism, the cyclin-CDK complex are also negatively regulated by the CDK inhibitors.

CDK inhibitors also referred to as CKI's act as important checkpoints responsible for the maintenance of cell cycle and proliferation arrest once the cells' development fate has reached. The CDK inhibitors either bind to CDK alone or to the CDK-cyclin complex and thus regulate CDK activity. There are two main classes of CKI's namely the Cip/Kip family which includes p21, p27 and p57 as its components and INK4 family includes p16, p14 and p20. The function of Cip/Kip family is mainly to inactivate cyclin D/CDK bond while INK4 family functions by blocking the cell cycle. The disruption of these check points is one of

the mechanism by which the tumor cell exhibits abnormal proliferation.

Oral leukoplakia is the most frequently encountered potentially malignant disorder of the oral mucosa with an increased potential for malignant transformation. The histopathological description oral leukoplakia generally described as epithelial dysplasia based on the architectural and cytological changes observed in the biopsy tissue (Warnakulasuriya et al 2008). While leukoplakia generally has a malignant transformation rate of less than 2%, those with histological evidence of epithelial reported dysplasia are to undergo malignant transformation in the range of 1.1% to 17.5% (Napier & Speight 2008). While it is generally accepted that the malignant transformation rate increases with severe grade of epithelial dysplasia, clear consensus can be drawn regarding the behavior of non-dysplastic lesions. In addition to clinical and histological changes, these lesions also show widespread genetic and molecular alterations. Molecular assessment of these lesions may thus help in early diagnosis and prevention of malignant transformation.

This paper mainly reviews the role of cell cycle inhibitors in oral leukoplakia and its potential for malignant transformation.

2. Cell cycle and its inhibitors

Cell division consists of two consecutive processes: mitosis, which is the process of nuclear division and; interphase which is the period between two mitoses. The interphase is characterized by G1, S and G2 phase. DNA replication occurs in S phase. The gap phase prior to the DNA replication is G1 phase where the cell is at preparatory stage and G2 phase occurs after DNA synthesis where the cell prepares for mitosis. Cells in the G0 phase includes non-growing, non-proliferating cells in the human body (Kumar et al 2015).

The cyclin dependent kinases (CDK's) regulate the cell cycle in the presence of cyclins. Expression of cyclins is cell cycle phase- dependent and is regulated transcriptionally, post-transcriptionally as well as translationally/

post-translationally (Lundberg & Weinberg 1999). Cyclins D1, D2 and D3 bind to CDK 4 and 6 in order to gain entry in G1 phase (Sherr 1994). Cyclin E binds to CDK2 to regulate progression from G1 to S phase (Ohtsubo et al 1995). Cyclin A interacts with CDK2 and this is required during G2 phase. For entry into M phase cyclin A interacts with CDK1 (Arellano & Moreno 1997). In somatic cells. movement through G1 and S phase is driven by the active form of cyclin D/CDK4,6 complex and subsequent phosphorylation of RB protein (Classon & Harlow 2002). The retinoblastoma (Rb) protein is a key switch at R point and phosphorylation results in structural conformation of proteins that initiate or inhibit cell physiological events (Todd et al 2002). Once Rb is phosphorylated, it partially releases critical transcription factor E2F-1 that turns on a series of genes coding for cyclin A and E that forms a complex with CDK2. The complex leads to complete release of E2F followed by the transcription of series of genes responsible for S-phase progression

and DNA synthesis (Bartek & Lukas 2001).

Cyclin D1 is a key component of cell cycle progression and interacts closely with the Rb protein. In the hypophosphorylated state, Rb protein acts as a tumor suppressor and contributes to cell cycle regulation at the G1 to S checkpoint by suppressing gene transcription that is required for entry into the S phase. The cell cycle is then arrested in G1. In response to mitogenic signals, CDK4 and CDK6 form a complex with their regulatory subunit, cyclin D1, which phosphorylates the Rb protein, reducing its ability to suppress gene transcription. Controlled phosphorylation deactivation of the Rb protein by the CDK4/6 complex is essential progression of the normal cell cycle (Shepperd & **MCarthur** 2013). In malignant cells, unrestricted CDK4/6 activity pathway can result from alterations in the expression of cyclindependent kinases and their regulatory mechanisms. This unhindered cell cycle stimulation yields a growth advantage and

uncontrolled cell proliferation (Shapiro 2006).

Cell cycle regulation is crucial in tumorigenesis and depends activities of CDK's which are positively regulated by mitogenic growth factors and negatively by CDK inhibitors (CKIs). These CKI's may either bind alone to CDK or to the CDK-cyclin complex, thereby regulating their activity. There are two main classes of CKIs based on their origin, structures and CDK specificities. They are the Cip/Kip family and INK4 family (Sherr and Roberts 1995). The Cip/Kip family encodes for p21, p27 and p57 which binds to both cyclin and CDK subunits thereby modulating the activities of cyclin D-, E-, A-, and B-CDK complexes. The INK4 gene family encodes p16, p15, p14 and p18 all of which bind to CDK 4 and 6 thus inhibiting their kinase activity by interfering with their association with cyclin D (Sherr and Roberts 1999). While the proteins of both the family share a conserved N-terminal domain, they diverge in the remainder of their sequence, suggesting that each of these proteins could have distinct

functions and regulation (Besson et al 2008).

2.1. Cyclin dependent kinases inhibitors

The activity of CDK's in cell cycle regulations is counteracted by their inhibitors known as cyclin dependent kinase inhibitors (CKIs) which function by either binding to CDK alone or to the CDK-cyclin complex. As already described, CKIs belong to two broad family of proteins namely Cip/Kip proteins and the INK4/ARF family.

The Cip/Kip proteins are intrinsically unstructured and they adopt specific tertiary conformations only after binding to other proteins (Lacy et al 2004). The member of this family namely p21, p27 and p57 function by inactivation of G1 cyclin- CDK complex and cyclin B-CDK1 complex. In addition, they can also modulate cell cycle progression via inhibition of components of the replication machinery. An important protein of this group, p21 was first reported to bind to proliferating cell nuclear antigen (PCNA) via its C-

terminus thereby blocking progressive DNA synthesis (Besson et al 2008). This protein is a critical downstream target in the p53-specific pathway of growth control, and can also be induced by p53 independent pathways in relation to terminal differentiation. The p21 gene which encodes this protein is present on chromosome 6p21.2 regions. In normal cells this protein predominantly exists in quaternary complexes with cyclins, CDKs and PCNA to inhibit the CDK activity and controls the G1 to S transition (Xiong et al 1993). The function of p21 includes differentiation of normal and transformed association with cells. terminal differentiation, senescence and apoptosis through p53 independent mechanisms (Agarwal et al 2008). Another member of this group, p27 was first identified as a CKI due to its ability to block the activity of cyclin E/CDK2 and cyclin A/CDK2 complex in cells arrested in G1 phase by transforming growth factor beta. lovastatin and contact inhibition. The expression of p27 is mainly regulated by ubiquitin-dependent proteolysis (Kudo et al 2005). They act mainly in G0 and early

G1 with the primary target being E-type cyclin/cdk2 complexes. Mitogenic growth factor signaling causes loss of p27 and their levels increase in response to differentiation signals (Slignerland et al 2000). By immunohistochemical staining, cells in the prickle cell and granular cell layers show strongly positive staining for p27 in their nuclei, but cells in the basal layer do not. Therefore, p27 is suggested to play an important role in cell cycle arrest in oral epithelial cells (Kudo et al 2005). The degree of p27 expression in various human malignancies is reported to be inversely correlated to malignant transformation of lesions (Kovesi and Szende 2006).

The INK4/ARF family are known to cause inactivation of G1 CDK (4 & 6) and the protein of this group are p15, p16, p18 and p14. The most important member is the p16 which is known to control cell cycle by inhibiting the ability of cyclin D-CDK4/6 complex to phosphorylate retinoblastoma protein (pRb). Thus, inactivation of pRb by phosphorylation causes p16 expression while hypophosphorylated active pRb can repress p16 expression (Li et al 1994). Inactivation of p16 was found in many cancers including head and neck squamous cell carcinoma and may occur by various mechanism such as point mutations, homozygous deletion and DNA methylation (Papadimitrakopoulou et al 1997)

3. Oral leukoplakia and its malignant transformation

Oral squamous cell carcinoma (OSCC) initiates in a multistep process in which normal cells are transformed into a pre-neoplastic cell followed by its malignant transformation. The majority of oral cancers are preceded by visible mucosal changes that have the propensity to develop into malignancy (Villa and Gohel 2014). These changes are currently grouped together and referred to as potentially malignant disorders (Van der Waal 2014). The concept of denoting these disorders as pre-cancerous is based on the fact that (i) they may undergo malignant changes over a period of time, (ii) they co-exist at the margins of clinically diagnosed carcinoma, (iii) they may share morphological and cytological changes observed in malignant lesions, but without frank invasion and (iv) they may demonstrate chromosomal, genomic and molecular alterations as observed in oral carcinoma (Warnakulasuriya et al 2007). The most common form of potentially malignant disorder leukoplakia which is defined as a white plaque of questionable risk having excluded other known diseases disorders that carry no risk of cancer (Warnakulasuriya et al 2007). The process of diagnosing leukoplakia begins with a provisional diagnosis which is made at the initial clinical examination when the lesion is not clearly diagnosed as any other entity. This is followed by a definite diagnosis which is made when any etiological cause other than tobacco/areca nut has been excluded and histopathology has not confirmed any other disorder (van der Wall and Axell 2002).

While leukoplakia is a clinical term, the histopathology of these lesions serve two important functions; to exclude any other definable lesion and to establish the degree of epithelial dysplasia if any (Van

der Waal et al 1997). The term epithelial dysplasia is applied when the tissue demonstrates architectural disturbances along with cytological atypia (Barnes et al 2005), and its presence serves as an important prognostic predictor of transformation. malignant The microscopic appearance oral leukoplakia varies from a simple hyperplasia characterized by acanthosis without cellular atypia to dysplasia. The histopathological features of epithelial dysplasia includes loss of polarity of the basal cells; presence of more than one layer having a basaloid appearance; dropshaped rete-ridges; increased nuclearcytoplasmic ratio: nuclear hyperchromatism; enlarged nucleoli; increased number of mitotic figures; abnormal mitosis; presence of mitotic figures in the superficial half of the epithelium; cellular and nuclear pleomorphism; irregular epithelial of stratification; loss intercellular adherence; keratinization of single cells or cell groups in the prickle cell layer. The dysplastic features can be graded as mild, moderate and severe based on

prominence of the features and may vary depending on the site of the lesion (Warnakulasuriya 2001).

The possible reason for the increase malignant transformation with increase in the severity of dysplasia could attributed to the accumulation of genomic and molecular alterations that may initiate cancer development. This, however, does not rule out the development of carcinoma from less severe dysplasia. Till date, there is no significant marker to understand and predict the potential of a lesion to undergo Genetic malignant transformation. mutation leading to potentially malignant lesions and oral malignancy mainly affects two set of genes namely the protooncogenes and tumor suppressor genes. The progress of a cell through various stages of cell cycle is tightly regulated by various mechanisms and disruption of these regulators may result in unrestrained proliferation of tumor cells. The CKIs are an important regulator of this complex process which undergo alteration thus leading to cell cycle deregulation. The identification and characterization of these cell cycle inhibitors could serve as an

important therapeutic target to prevent the malignant transformation of oral leukoplakia and thus provide a better prognosis.

oral leukoplakia and OSCC. The studies pertaining to the role of various CKIs in oral leukoplakia and its malignant transformation are highlighted in the table.

4. CKIs in oral leukoplakia

Over the years, scientific data has been amassed to assess the role of CKIs in

Sl.	Author and year	Type of CKIs studied	Results &	Conclusion
No		and the lesion	interpretation	
	Kovesi and Szende (2006)			These gene products could be useful tool for a precise prognosis of oral leukoplakia and its malignant transformation
			leukoplakia which	
			could be due to	
			defense	
			mechanism against	
			malignant	

			transformation	
2	Shintani S et al (2002)	p16, p12 and p27 in normal, oral epithelial dysplasia and OSCC	- Decreased expression of cell cycle inhibitors in dysplasia and OSCC than in normal controlsSignificant loss of expression in OSCC than in epithelial dysplasia	Loss of CKI expression may be associated with poor prognosis and contribute to multistep nature of oral carcinogenesis
3	Visioli et al (2012)	p21 expression between non-dysplastic leukoplakia and dysplastic leukoplakia	Overexpression of p21 in both dysplastic and non-dysplastic leukoplakia than in normal controls was evident. However similar expression profile was seen in both forms of leukoplakia.	Increased expression could represent an attempt to control cell proliferation, which is possibly overcome by other factors which stimulate carcinogenesis and overload the inhibitory function of p21
4	Papadimitrakopoulou	p16 expression in oral	47% of patients	The finding

	et al 1997)	leukoplakia	with oral	supports the
			premalignant	important role
			lesions	of p16 gene in
			demonstrated lack	head and neck
			of p16 expression.	tumorigenesis
			8 of these patients	
			developed	
			carcinoma and	
			among them, 5	
			patients showed	
			complete loss of	
			p16 expression	
5	Agarwal et al (1998)	p21 expression in	The result showed	Heterogeneity
		normal, oral leukoplakia	negligible staining	in p21
		and OSCC patients with	in normal controls,	expression was
		clinical correlation	60% of	observed in oral
			leukoplakia and	SCCs and in
			68% of OSCC	premalignant
			showed positive	lesions,
			expression of p21.	suggesting that
			Expression	alterations in
			decreased with	p21 expression
			increased grades of	are an early
			OSCC.	event in oral
				oncogenesis.
6	Kresty et al (2008)	p16 and p14 expression	Loss of p16 and	May contribute
		in proliferative	p14 expression	to the
		verrucous leukoplakia		aggressiveness
				and high rates

				of malignant
				transformation.
7	Buajeeb et al (2009)	p16 expression in oral	No significant	p16 could not
		leukoplakia with and	difference in p16	be used as a
		without dysplasia and	expression among	reliable marker
		OSCC	the study groups.	for oral mucosal
				dysplasia and
				malignant
				transformation
8	Bradley et al (2006)	p16 expression in oral	Loss of p16	Decreased
		epithelial dysplasia	expression in 36%	expression of
			of non-dysplasia,	p16 in
			39% mild	dysplastic
			dysplasia and 66%	lesions, may
			of moderate/severe	reflect the
			dysplasia	biologic events
				involving loss
				of p16 gene
				function in the
				pathogenesis of
				oral cancer.
				The study also
				indicates that
				p16 is not useful
				in
				differentiating
				dysplastic and
				non-dysplastic
				oral lesions

		T	T	
				though
				decreased p16
				expression with
				increasing
				severity of
				dysplasia was
				observed.
9	Nasser et al (2011)	p16, p53, cyclin D1	Loss of p16	Loss of p16 is
		expression in non-	expression was	an early and
		dysplastic leukoplakia,	seen in 32.4% of	common event
		dysplastic leukoplakia	leukoplakia	in oral
		and OSCC for	without dysplasia.	carcinogenesis.
		comparison	The frequency of	A combined
			loss increased	alteration of
			linearly with the	various proteins
			histopathological	could serve as a
			state	biomarker to
				define high risk
				leukoplakia
				patients.
				Lesions that do
				not show these
				alterations can
				be considered
				harmless.
10	Ramasubramanian A	p27, p63 and cyclin D1	Significantly	The findings
	(2013)	in varying grades of	increased	may serve as a
		epithelial dysplasia	expression of	prognostic
			cyclin D1 and p63	marker for any

			with moderately	preceding
			-	
			significant	malignant
			decrease in p27	transformation
			expression with	and in oral
			increasing grades	cancer
			of dysplasia was	progression
			observed.	
11	Tsuzuki et al (2003)	p27, cyclin D1, PCNA	p27 was expressed	The study
		expression in oral	in 34.8% of	suggests that the
		leukoplakia and	leukoplakia with	abundance of
		apoptotic index.	hyperplasia, 55.4%	p27 in oral
			in leukoplakia with	leukoplakia may
			dysplasia and	inhibit cell
			31.1% in SCC	proliferation
			(43.3% in the early	and lead
			stage of SCC,	premalignant
			25.9% in the	tumor cells to
			advanced stage of	apoptosis, and
			SCC). The level of	thus is
			p27 expression	concerned with
			showed a peak in	prevention of
			dysplasia and	tumor
			decreased in SCC	progression.

From the data available it is evident that there are contradictory results obtained pertaining to the exact role of CKIs. This could be attributed to inadequate sample size, lack of

standardization with respect to the staining pattern, and possible variation in the clinical presentation of the lesion that may influence sample selection. The genetic changes evident in oral

carcinogenesis are complex and there is a dynamic interplay of these changes which might influence the overall pathogenesis of oral cancer and hence it is difficult to attribute the malignant transformation to a single factor. However the following interpretations can be derived from the available data in the literature: a) loss of cell cycle inhibitor proteins are associated with poor prognosis and malignant transformation of oral leukoplakia b) it is necessary to analyze a panel of CKIs rather than an isolated protein to derive appropriate conclusions; and these studies invite exciting opportunities to gain insight into the genetic events that drive the carcinogenesis process and the effects of chemopreventive interventions possible modification of the changes that define the malignant phenotype.

5. Conclusion

Cell cycle and its inhibitors play a significant role in the malignant transformation of potentially malignant disorders and oral carcinogenesis. But modulation of cell cycle has also been known to contribute to chemotherapy resistance. It is thus prudent to understand the role of CKIs to predict the biological behavior of a suspicious lesion as well as implement relevant therapeutic to modalities. A wide range of anti-cancer drugs directed at CKIs are at different stages of clinical trials and it is only a matter of time before they are available for clinical use.

6. References

Agarwal S, Mathur M, Shukla NK, Ralhana R (1998). Expression of cyclin dependent kinase inhibitor p21waf1/cip1 in premalignant and malignant oral lesions: relationship with p53 status. Oral Oncology; 34:353-360

Arellano M, Moreno S (1997) Regulation of CDK/cyclin complexes during the cell cycle. Int. J. Biochem. Cell Biol; 29:559-573.

Barnes L, Eveson JW, Reichart P, Sidransky. WHO classification of tumors. Pathology and genetics. Head and neck tumors. International agency for research on cancer (IARC). Lyon, France: IARFC press 2005

Bartek J, Lukas J (2001). Pathways governing G1/S transition and their response to DNA damage. FEBS Lett; 490: 117–122.

Besson A, Dowdy SF, Roberts JM (2008). CDK inhibitors: cell cycle regulators and beyond. Developmental cell review. 159-169

Bradley KT, Budnick SD, Logani S (2006). Immunohistochemical detection of p16INK4a in dysplastic lesions of the oral cavity. Mod Pathol; 19(10):1310-1316.

Cheng T (2004). Cell cycle inhibitors in normal and tumor stem cells. Oncogene; 23:7256-7266

Classon M, Harlow E (2002). The retinoblastoma tumor suppressor in development and cancer. Nat. Rev. Cancer; 2:910–917.

Dickson MA, Schwartz GK (2009). Development of cell cycle inhibitors for cancer therapy. Current Oncol; 16(2):36-43

Kaldis P, Russo AA, Chou HS, Pavletich NP, Solomon MJ (1998). Human and yeast CDK- activating kinases (CAKs) display distinct substrate specificities. Mol Biol Cell; 9:2545-2560

Kovesi G, Szende B (2006). Prognostic value of cyclin D1 p27 and p63 in oral leukoplakia. J Oral Pathol Med; 35:274-277.

Kresty LA, Mallery SR, Knobloch TJ, Li J, Lloyd M, Casto BC, Weghorst CM (2008). Frequent Alterations of p16INK4a and p14ARF in Oral Proliferative Verrucous Leukoplakia. Cancer Epidemiol Biomarkers Prev; 17:3179-3187

Kudo Y, Kitajima S, Ogawa I, Miyauchi M, Takata T (2005). Down-regulation of Cdk inhibitor p27 in oral squamous cell carcinoma Oral Oncology; 41:105–116

Kumar, Abbas, Aster (2015). Robbins and Cotran Pathological basis of disease 9th edtn. Elsevier Publications (South Asia)

Lacy ER, Filippov I, Lewis WS, Otieno S, Xiao L, Weiss S, Hengst L,Kriwacki RW (2004). p27 binds cyclin-CDK complexes through a sequential mechanism involving binding-induced protein folding. Nat. Struct. Mol. Biol. 11, 358–364.

Li Y, Nichols MA, Shay JW, Xiong Y (1994). Transcriptional repression of the D-type cyclin dependent kinase inhibitor p16 by the retinoblastoma susceptibility gene product pRb. Cancer Res; 54:396-403

Lundberg AS, Weinberg RA (1999). Control of the cell cycle and apoptosis. Eur J Cancer 35:1886-1894.

Morgan DO (1995). Principles of CDK regulation. Nature; 374:131-134.

Napier SS, Speight PM (2008). Natural history of potentially malignant oral lesions and conditions: an overview of the literature. J Oral Pathol Med; 37:1-10

Nasser W, Flechtenmacher C, Holzinger D, Hofele C, Bosch FX (2011). Aberrant expression of p53, p16^{INK4a} and Ki-67 as basic biomarker for malignant

progression of oral leukoplakias. J Oral Pathol Med; 40:629-635.

Ohtsubo M, Theodoras AM, Schumacher J, Roberts JM, Pagano M (1995). Human cyclin E, a nuclear protein essential for the G1-to-S phase transition. Mol Cell Biol; 15:2612-2624.

R. Todd, PW. Hinds, K. Munger, AK. Rustgi, OG. Opitz, Y. Suliman, DT. Wong (2002). Cell cycle dysregulation in oral cancer Crit Rev Oral Biol Med; 13(1):51-61

Ramasubramanian A, Ramani P, Sherlin HJ, Premkumar P, Natesan A, Thiruvengadam C (2013). Immunohistochemical evaluation of oral epithelial dysplasia using cyclin-D1, p27 and p63 expression as predictors of malignant transformation. J Nat Sci Biol Med; 4:349-58.

Shapiro GI (2006). Cyclindependent kinase pathways as targets for cancer treatment. J Clin Oncol. 2006;24(11):1770-1783.

Sheppard KE, McArthur GA (2013). The cell-cycle regulator CDK4: an emerging therapeutic target in melanoma. Clin Cancer Res. 2013;19(19):5320-5328.

Sherr CJ (1994) G1 phase progression: cycling on cue. Cell; 79:551-555.

Sherr CJ, Roberts JM (1995) Inhibitors of mammalian G1 cyclindependent kinases. Genes Dev. **9**, 1149-1163.

Sherr CJ, Roberts JM (1999). CDK inhibitors: positive and negative regulators of G1-phase progression. Genes Dev; 13:1501–1512.

Shintani S, Mihara M, Nakahara Y, Kiyota A, Ueyama Y, Matsumura T, Wong DT (2002). Expression of cell cycle control proteins in normal epithelium, premalignant and malignant lesions of oral cavity. Oral Oncol; 38(3):235-43.

Slingerland J, Pagano M (2000). Regulation of the Cdk Inhibitor p27 and Its Deregulation in Cancer. Journal of cellular physiology 183:10–17

Tsuzuki H, Fujieda S, Sunaga H, Narita N, Tokuriki M, Saito H (2003). Expression of p27 and apoptosis in oral leukoplakia. Anticancer Res; 23:1265-70.

Papadimitrakopoulou V, Izzo J, Lippman SM, Lee JS, Fan YH, Clayman G, Ro JY, Hittelman WN, Lotan R, Hong WK, Mao L (1997). Frequent inactivation of p16INK4a in oral premalignant lesions. Oncogene; 14:1799-1803 Van der Waal I (2014). Oral potentially malignant disorders: Is malignant transformation predictable and preventable? Med Oral Patol Oral Cir Bucal; 19:e386-e390

Van der Waal I, Schepman KP, Van der Meij EH, Smeele LE (1997). Oral Leukoplakia: a clinico-pathological Review. Oral Oncol; 33:291-301

Vermeulen K, Van Bockstaele DR, Berneman ZM (2003). The cell cycle: a review of regulation, deregulation and therapeutic targets in cancer. Cell prolif; 36: 131-149

Villa A, Gohel A (2014). Oral potentially malignant disorders in a large population. J Appl Oral Sci; 22:473-476

Visioli F, Lauxen IS, Filho MS, Rados PV (2012). Expression of the cell cycle regulation proteins p53 and p21waf1 in different types of non-dysplastic leukoplakias. J Appl Oral Sci; 20(3):369-75

Warnakulasuriya S (2001). Histological grading of oral epithelial dysplasia: revisited. J Pathol; 194:294-297

Warnakulasuriya S, Johnson NW, Van der Waal I (2007). Nomenclature and classification of potentially malignant disorders of the oral mucosa. J Oral Pathol Med; 36:575–80

Warnakulasuriya S, Reibel J, Bouquot J, Dabelsteen E (2008). Oral epithelial dysplasia classification systems: predictive value, utility, weaknesses and scope for improvement. J Oral Pathol Med; 37: 127-133 Xiong Y, Hannon GJ, Zhang H, Casso D, Kobayashi R, Beach D. p21 is a universal inhibitor of cyclin kinases. Nature 1993; 366:701-704.